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U.S. DEPARTMENT OF COMMERCE PATENT AND TRADEMARK OFFICE

ATTORNEY'S DOCKET NUMBER

TRANSMITTAL LETTER TO THE UNITED STATES  
DESIGNATED/ELECTED OFFICE (DO/EO/US)  
CONCERNING A FILING UNDER 35 U.S.C. 371

1659/3

U.S. APPLICATION NO. (IF KNOWN, SEE 37 CFR 1.5)

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INTERNATIONAL APPLICATION NO.

129910024

INTERNATIONAL FILING DATE

8 MAY 1997

PRIORITY DATE CLAIMED

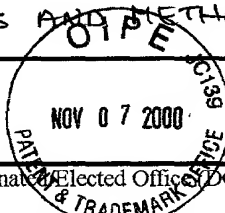
8 MAY 1998

TITLE OF INVENTION

CARBOXYLIC CATIONITES AND METHODS OF MANUFACTURE

APPLICANT(S) FOR DO/EO/US

KARINA ROZHETSKY



Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:

1. ☒ This is a **FIRST** submission of items concerning a filing under 35 U.S.C. 371.
2. ☐ This is a **SECOND** or **SUBSEQUENT** submission of items concerning a filing under 35 U.S.C. 371.
3. ☒ This express request to begin national examination procedures (35 U.S.C. 371(f)) at any time rather than delay examination until the expiration of the applicable time limit set in 35 U.S.C. 371(b) and PCT Articles 22 and 39(1).
4. ☒ A proper Demand for International Preliminary Examination was made by the 19th month from the earliest claimed priority date.
5. ☒ A copy of the International Application as filed (35 U.S.C. 371 (c) (2))
  - a. ☒ is transmitted herewith (required only if not transmitted by the International Bureau).
  - b. ☐ has been transmitted by the International Bureau.
  - c. ☐ is not required, as the application was filed in the United States Receiving Office (RO/US).
6. ☐ A translation of the International Application into English (35 U.S.C. 371(c)(2)).
7. ☒ A copy of the International Search Report (PCT/ISA/210).
8. ☐ Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371 (c)(3))
  - a. ☐ are transmitted herewith (required only if not transmitted by the International Bureau).
  - b. ☐ have been transmitted by the International Bureau.
  - c. ☐ have not been made; however, the time limit for making such amendments has NOT expired.
  - d. ☐ have not been made and will not be made.
9. ☐ A translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)).
10. ☒ An oath or declaration of the inventor(s) (35 U.S.C. 371 (c)(4)).
11. ☒ A copy of the International Preliminary Examination Report (PCT/IPEA/409).
12. ☐ A translation of the annexes to the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371 (c)(5)).

## Items 13 to 18 below concern document(s) or information included:

13. ☐ An Information Disclosure Statement under 37 CFR 1.97 and 1.98.
14. ☒ An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.
15. ☐ A **FIRST** preliminary amendment.  
A **SECOND** or **SUBSEQUENT** preliminary amendment.
16. ☐ A substitute specification.
17. ☐ A change of power of attorney and/or address letter.
18. ☐ Other items or information:

09/674825

1199/00291

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529 Rec'd PCT/PTC 07 NOV 2000

20. The following fees are submitted:

CALCULATIONS PTO USE ONLY

## BASIC NATIONAL FEE (37 CFR 1.492 (a) (1) - (5)):

- ☐ Search Report has been prepared by the EPO or JPO . . . . . \$ 840
- ☐ International preliminary examination fee paid to USPTO (37 CFR 1.482) . . . . . \$ 670
- ☐ No international preliminary examination fee paid to USPTO (37 CFR 1.482) but international search fee paid to USPTO (37 CFR 1.445(a)(2)) . . . . . \$ 690
- ☐ Neither international preliminary examination fee (37 CFR 1.482) nor international search fee (37 CFR 1.445(a)(2)) paid to USPTO . . . . . \$ 970
- ☒ International preliminary examination fee paid to USPTO (37 CFR 1.482) and all claims satisfied provisions of PCT Article 33(2)-(4) . . . . . \$ 96

ENTER APPROPRIATE BASIC FEE AMOUNT =

96

Surcharge of \$130.00 for furnishing the oath or declaration later than months from the earliest claimed priority date (37 CFR 1.492 (e)). ☐ 20 ☐ 30

| CLAIMS             | NUMBER FILED | NUMBER EXTRA | RATE   |
|--------------------|--------------|--------------|--------|
| Total claims       | 44 - 20 =    | 24           | x \$15 |
| Independent claims | 4 - 3 =      | 1            | x \$78 |

432

78

Multiple Dependent Claims (check if applicable). ☐

TOTAL OF ABOVE CALCULATIONS =

606

Reduction of 1/2 for filing by small entity, if applicable. Verified Small Entity Statement must also be filed (Note 37 CFR 1.9, 1.27, 1.28) (check if applicable). ☒

SUBTOTAL =

303

Processing fee of \$130.00 for furnishing the English translation later than months from the earliest claimed priority date (37 CFR 1.492 (f)). ☐ 20 ☐ 30 +

TOTAL NATIONAL FEE =

Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31) (check if applicable). ☒

40

TOTAL FEES ENCLOSED =

343

|               |    |
|---------------|----|
| Amount to be: | \$ |
| refunded      |    |
| charged       | \$ |

- ☐ A check in the amount of \_\_\_\_\_ to cover the above fees is enclosed.
- ☒ Please charge my Deposit Account No. 06-2140 in the amount of & \$ 343 to cover the above fees. A duplicate copy of this sheet is enclosed.
- ☐ The Commissioner is hereby authorized to charge any fees which may be required, or credit any overpayment to Deposit Account No. \_\_\_\_\_ A duplicate copy of this sheet is enclosed.

NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137(a) or (b)) must be filed and granted to restore the application to pending status.

SEND ALL CORRESPONDENCE TO:

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SIGNATURE

MARK M. FRIEDMAN

NAME

33,883

REGISTRATION NUMBER

DATE

## SMALL BUSINESS CONCERN - NEW APPLICATION

Attorney Docket No. 1659/3

## IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In RE Application of: KARINA ROZHETSKY

Filed Concurrently Herewith

For: CARBOXYLIC CATIONITES AND METHODS OF MANUFACTUREVERIFIED STATEMENT UNDER 37 CFR 1.27  
CLAIMING STATUS AS A SMALL ENTITY

To The Commissioner of Patents and Trademarks:

I hereby declare that:

I am the owner of, or an official empowered to act on behalf of, the small business concern identified below.

Name of Concern: POLYGRAN LTD.Address : HAHISTADRU BLVD. 90, 32960 JHAQFA, ISRAEL

The small business concern identified above, together with its affiliates, employs fewer than 500 persons and qualifies as a small business concern as defined in 37 CFR 1.9(d) for purposes of claiming reduced fees under 35 USC § 41(a) and § 41(b) to the Patent and Trademark Office with regard to the above-entitled invention described in the specification filed herewith.

Rights under contract or law have been conveyed and remain with the small business concern identified above with regard to the above entitled invention.

If the rights held by the small business concern are exclusive, each other party having rights to the invention is listed below, and no rights to the invention are held by any party who could not be classified as a small entity under 37 CFR 1.9(f), namely any person who could not be classified as an independent inventor under 37 CFR 1.9(c) if the person had made the invention, or any concern which would not qualify as a small business concern under 37 CFR 1.9(d) or a nonprofit organization under 37 CFR 1.9(e).

Full Name (Party 1) : NONE

Address : \_\_\_\_\_

Status : ☒ Individual ☐ Small Business Concern ☐ Nonprofit Organization

Full Name (Party 2) : \_\_\_\_\_

Address : \_\_\_\_\_

Status : ☐ Individual ☐ Small Business Concern ☐ Nonprofit Organization

I acknowledge the duty under 37 CFR 1.28(b) to file, in this application, notification of any change in status resulting in loss of entitlement to small entity status prior to paying, or at the time of paying, the issue fee due after the date on which status as a small entity is no longer appropriate.

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application and any patent issuing thereon.

Rozhetsky  
Name of Person Signing

Perry  
Signature

3.11.00  
Date

Capacity of Person Signing: 1Address of Person Signing: Peor Street 32, 32492 Haifa, Israel

529 Rec'd PCT/PTC 07 NOV 2000

Carboxylic Cationites and Methods Of ManufactureFIELD AND BACKGROUND OF THE INVENTION

The present invention is of carboxylic cationites with improved sorption capacity,  
5 particularly for high molecular weight substances such as proteins, polysaccharides and other  
macromolecules as well as for improved methods for manufacturing the carboxylic cationites.

Ion exchange is one of the most common procedures for biomolecules isolation and  
purification, and there are many ion exchanging supports for bio-chromatography. The ion-  
exchange chromatography technique is based on the ability to separate charged substances by  
10 'adherence' to resins carrying the opposite charge. The ion groups of exchange are bound  
through a covalent bond to the absorbing matrix.

Many currently available background art sorbent materials for ion-exchange  
chromatography are based on various polysaccharides such as those known under the trade  
names Sepharose™, Sephadex™ and so forth. Unfortunately these sorbent materials have only  
15 limited sorption ability and are not able to fully separate macromolecules, such as proteins,  
from complex mixtures in the presence of mineral salts. Therefore, such background art  
sorbent materials are useful only after the stage of removing mineral salts from the mixture, or  
"desalting". In addition, the chemical stability and thermostability of these materials is low,  
and accordingly, they are useful only for a limited number of regeneration cycles.

20 Carboxylic cationites have a number of advantages over such polysaccharide-based  
sorbents, and as such are known and widely used for sorption and desorption of various  
macromolecules such as proteins. One of the advantages of these cationites is that they do not  
denature the proteins and enzymes during the process of separation. Unfortunately, those  
carboxylic cationites which are known in the background art largely feature a macroporous  
25 structure which has a high flow rate and a low exchange capacity, with limited sorption of  
macromolecules such as proteins. This structure limits the usefulness of carboxylic cationites  
for ion exchange chromatography with such macromolecules. The structure of the carboxylic  
cationites is in turn a result of the method employed for manufacturing these cationites.

US Patent No. 4,128,032 discloses one such background art method of manufacture  
30 for carboxylic cationites. This process involves the co-polymerization of methacrylic or  
acrylic acids with a suitable crosslinking agent, such as hexahydro-1,3,5 -triacryloyl triazine  
(HTA), N,N'-methylenediacrylamide (MDAA), N,N'-ethylenedimethacrylamide (EDMA), or

N,N'-hexamethylenedimethacrylamide (HMDMA) with an initiator of radical polymerization. The resulting reaction mixture is dispersed in high viscosity liquid, either liquid polyethylsiloxane or polymethylsiloxane, as the dispersing medium.

Unfortunately, the carboxylic cationites prepared according to this background art process do not possess sufficient sorption capacity required for the effective separation of macromolecules such as proteins and enzymes. This deficiency may be caused by the phenomena of absorption and surface tension on the phase interface of the reaction mixture and liquid polysiloxane. This tension may decrease the size of pores formed on the outside surface of the granules of carboxylic cationites. Furthermore, such a process for manufacturing the cationites also has the disadvantage of requiring an organic solvent to wash out the dispersion medium, which is a potential safety hazard and which increases the expense of manufacture.

A further disadvantage of this background art method is the migration of components of the reaction mixture (comonomers and acetic acid) into the dispersion medium, which occurs during the stage of copolymerization. Such migration increases the difficulty of preparing carboxylic cationite granules with reproducible structural properties. Thus, the disclosed background art method has a number of disadvantages.

There is thus a need for, and it would be useful to have, a method for manufacturing carboxylic cationites which requires little or no organic solvent in the dispersion medium, and which results in the production of cationites with reproducible structural properties and efficient sorption of macromolecules such as proteins.

### Summary of the Invention

The present invention is of a method for manufacturing carboxylic cationites which optionally does not include a dispersion stage. If the dispersion stage is performed, preferably the dispersion medium includes an aqueous solution of an inorganic salt. Optionally, if two dispersion stages are performed, one stage is performed with alcohol or a mixture of alcohols.

Various co-polymerization or pre-polymerization solutions are possible. For example, one such solution is a mixture of an aqueous solution and an organic solvent. Preferably, the organic solvent is selected from the group consisting of dimethylformamide, monobutyl glycolate, polyethylene glycol and an alcohol. More preferably, the alcohol is selected from the group consisting of butyl alcohol, octyl alcohol and decyl alcohol. Alternatively, the co-

polymerization or pre-polymerization solution is a mixture of an organic solvent and acetic acid. More preferably, the range of concentration of acetic acid is from about 20 percent to about 50 percent volume per volume of the solution for such a mixture. Another preferred solution features an aqueous solution of acetic acid, more preferably in a range of from about  
5 60 percent to about 100 percent, weight per weight.

According to optional but preferred embodiments of the present invention, the dispersion stage is performed in two stages, in which one stage includes dispersion in an aqueous medium containing an inorganic salt, while the other stage includes dispersion in alcohol. The order of these stages preferably depends upon the concentration of the co-  
10 monomers, as described in greater detail below.

According to other optional but preferred embodiments of the present invention, no prepolymerization stage is performed, such that the co-monomers undergo co-polymerization directly, without first being incubated during the prepolymerization stage.

Monovinyl monomers and a cross-linking agent are employed as co-monomers, as  
15 described in further detail below. The co-monomers are dissolved in the selected solvent for co-polymerization or pre-polymerization, as described previously. A free radical initiator is then added, also as described in further detail below.

The resultant carboxylic cationites are obtained in one of three forms: a swollen block after co-polymerization in the selected solution, if no dispersion stage is performed; and, if a  
20 pre-polymerization stage and a dispersion stage is performed, depending upon the dispersion conditions, the resultant structures can have an irregular or regular form, and can be particles or spherical granules. The typical mean size of the resulting particulate product is in a range of from about 10 to about 200 microns. The structure of the resultant product typically features pores of at least about 0.05 microns (500 Angstrom) in diameter, or even greater sizes. By  
25 contrast, carboxylic cationites produced according to the background art method disclosed in U.S. Patent No. 4,120,831, in which the pre-polymerization solution contains from 5 percent to 50 percent acetic acid in water, and the dispersion medium is a liquid polysiloxane organic solvent, have pores of only 0.02 microns (200 Angstrom) in diameter. The properties of the  
30 cationites produced according to the method of the present invention result in improved sorption capacity for macromolecules such as proteins, and is suitable for isolation and purification of a wide range of proteins.

According to the present invention, there is provided a method for producing a carboxyl cationite, comprising the steps of: (a) dissolving a monovinyl monomer and a cross-linking agent as co-monomers in a first solvent to form a first mixture, the first solvent being selected from the group consisting of a mixture of an organic solvent and water, a mixture of an organic solvent and acetic acid, and an aqueous solution of acetic acid wherein an amount of the acetic acid is in a range of from about 60% to about 100%, volume per volume; and (b) adding an initiator of radical polymerization to the first mixture to form a swollen block of the carboxyl cationite, thereby obviating the need for dispersing the carboxyl cationite in a dispersing medium.

According to still another embodiment of the present invention, there is provided a method for forming particulate matter of carboxylic cationites, the method comprising the steps of: (a) dissolving a monovinyl monomer and a cross-linking agent as co-monomers in a first solvent to form a first mixture; (b) adding an initiator of radical polymerization to the first mixture to form a pre-polymer; (c) dispersing the pre-polymer in a solution of alcohol to form a dispersion mixture; and (d) mixing the dispersion mixture with an aqueous solution of an inorganic salt to form the particulate matter.

According to yet another embodiment of the present invention, there is provided a method for forming particulate matter of a carboxylic cationite, the method comprising the steps of: (a) dissolving a monovinyl monomer and a cross-linking agent as co-monomers in a first solvent to form a first mixture; (b) adding an initiator of radical polymerization to the first mixture to form a pre-polymer; and (c) dispersing the pre-polymer in an aqueous solution of an inorganic salt to form the particulate matter.

Hereinafter, the term "macromolecule" refers to at least one such molecule, and optionally a complex of a plurality of such molecules, which could be optionally covalently or non-covalently linked, including but not limited to a protein, polysaccharide, oligopeptide, and an oligonucleotide. Preferably, such a macromolecule has a molecular weight in the range of from about 60,000 Daltons to about 900,000 Daltons.

Hereinafter, the term "aqueous solution" includes water alone as the solution.

Hereinafter, the term "particulate matter" includes, but is not limited to, particles and granules, whether irregular or regular in shape.

#### Detailed Description of Preferred Embodiments

The present invention is of a method for manufacturing carboxylic cationites with three different structures, according to the embodiment of the method of the present invention which is used.

According to the first embodiment, co-monomers and an initiator are incubated in a solution which is preferably selected from the group consisting of a mixture of water and an organic solvent, a mixture of an organic solvent and acetic acid, and an aqueous solution of acetic acid wherein the amount of the acetic acid is in a range of from about 60 percent to about 100 percent, weight per weight. Most preferably, the solution is the aqueous solution of acetic acid wherein the amount of the acetic acid is in a range of from about 60 percent to about 100 percent, weight per weight. Preferably, the organic solvent is selected from the group consisting of dimethylformamide, monobutyl glycolate, polyethylene glycol and an alcohol. More preferably, the alcohol is selected from the group consisting of butyl alcohol, octyl alcohol and decyl alcohol. Alternatively, the co-polymerization solution is a mixture of an organic solvent and acetic acid. More preferably, the range of concentration of acetic acid is from about 20 percent to about 50 percent volume percent of the total volume of the solution. The co-monomers undergo co-polymerization directly, without the performance of a pre-polymerization stage or a dispersion stage, resulting in the preparation of a swollen block of cationites. For the preparation of the swollen block, the amount of the co-monomers in the selected co-polymerization solution is more preferably in the range of from about 10 percent to about 30 percent weight per weight.

According to a second embodiment of the method of the present invention, irregular particles are obtained from either a high concentration or a low concentration co-monomer solution. For the third embodiment, spherical granules are obtained from such low concentration or high concentration co-monomer solutions.

For the low concentration co-monomer solution, preferably the concentration of co-monomer is in the range of from about 20 percent to about 30 percent weight per weight. For the high concentration co-monomer solution, preferably the concentration of co-monomer is in the range of from about 50 percent to about 70 percent weight per weight. The preparation of the cationites includes two stages: a pre-polymerization stage, and a dispersion stage.

For the pre-polymerization stage, preferably the co-monomers are incubated with the initiator in a solution which is an aqueous solution of acetic acid. More preferably, the amount of acetic acid is in a range of from about 60 percent to about 100 percent, or



alternatively in a range of from about 5 percent to about 15 percent, weight per weight. A mixture of an organic solvent and acetic acid is alternatively and optionally used, as described previously.

The conditions for the dispersion stage(s) preferably differ according to whether a low concentration or a high concentration solution of co-monomers is used for the pre-polymerization stage. If a low concentration solution is used, then preferably the pre-polymer is placed first into a solution of alcohol as the dispersion medium to form a dispersed mixture. More preferably, the ratio of pre-polymer to alcohol is in a range of from about 2.5:1 to about 5:1 to form irregular particles, and is 1:1 to form spherical granules. Next, the dispersed mixture is preferably placed in an aqueous solution of an inorganic salt, more preferably an aqueous solution of sodium sulfate, and most preferably a 20 percent weight per weight aqueous solution of sodium sulfate. The ratio of dispersion mixture to aqueous dispersion solution is preferably in a range of from about 1:4 to about 1:5.

If a high concentration solution is used, then preferably the pre-polymer is placed first into an aqueous solution of an inorganic salt to form the dispersion mixture. The aqueous solution is more preferably an aqueous solution of sodium sulfate, and most preferably a 20 percent weight per weight aqueous solution of sodium sulfate. The ratio of pre-polymer to aqueous dispersion solution is preferably in a range of from about 1:4 to about 1:5.

Optionally and more preferably, a solution of alcohol is added to the dispersion mixture.

More preferably, the ratio of dispersed mixture to alcohol is in a range of from about 2:1 to about 1:1. If the second dispersion stage in alcohol is not performed, then irregular particles are formed. If both stages are performed, then spherical granules are formed.

For all three embodiments of the method of the present invention, monovinyl monomers and a cross-linking agent are employed as co-monomers. Preferred monovinyl monomers are selected from the group consisting of methacrylic acids, acrylic acids and methacryloylaminocarbonic acids. Examples of such monovinyl monomers include, but are not limited to, methacrylic acid, acrylic acid, methacryloylcaproic acid, methacryloylaminobenzoic acid, methacryloylglycine and methacryloylaminocarbonic acid. The cross-linking agent is preferably a long chain cross-linking agent of at least two repeated carbon groups in the chain, and more preferably of from about 2 to about 10 repeated carbon groups. Most preferably, the cross-linking agent is a polyvinylmonomer selected from the group consisting of hexahydro-1,3,5 -triacryloyltriazine (HTA), N,N'-methylenediacrylamide

(MDAA), N,N'-ethylenedimethacrylamide (EDMA), N,N'-hexamethylenedimethacrylamide (HMDMA), triallylisocyanurate (TAIC), *p*-phenylenedimethacrylamide (*p*-PHDMA), benzidinedimethacrylamide (BDMA) and so forth. Most preferably, the polyvinylmonomer is selected from the group consisting of HTA, EDMA, and TAIC. Also most preferably, the weight ratio of the monovinyl monomer to the polyvinyl-containing cross-linking agent is in the weight ratio of from about 8:1 to about 16:1.

The co-monomers are dissolved in the selected solvent, in which the initiator is also dissolved, for a prepolymerization stage, or else the co-polymerization stage if no prepolymerization stage is performed. The initiator is preferably a free radical initiator selected from the group consisting of ammonium persulfate, a 1,1' azobis (cyclohexanecarbonitrile), and a suitable redox system such as ammonium persulfate - sodium methabisulfite.

The pre-polymerization or co-polymerization stage is preferably performed at a temperature in a range of from about 20 to about 25 °C if the initiator is a redox system, or alternatively at a temperature in a range of from about 50 to about 60 °C if the initiator is ammonium persulfate or a 1,1' azobis (cyclohexanecarbonitrile).

The form and structure of the resultant carboxylic cationites depends upon the conditions of manufacture. If no prepolymerization or dispersion stages are performed, such that the co-monomers are dissolved with the initiator in the selected solution for co-polymerization, then the resultant carboxylic cationites are obtained as a swollen block. The block is then preferably crushed and washed in an aqueous solution. More preferably, the block is washed first with water, then with an aqueous solution of from about 0.5 N to about 1 N sodium hydroxide of pH 8, then with a 1 N aqueous solution of hydrochloric acid, and finally again by water. After these washing steps, the copolymer is fractionated in the wet state. Preferably, the fraction of particles of 100-200 micron diameter is used in the chromatographic column.

Alternatively, if a pre-polymerization stage is performed, then the final form and structure depends upon the dispersion medium. For a low concentration solution of co-monomers in the range of from about 20 percent to about 30 percent weight percent, if the pre-polymer is dispersed first in alcohol at a concentration of 1:1 alcohol to pre-polymer, then spherical granules are obtained. Otherwise, if the ratio of alcohol to pre-polymer is less than 1:1 volume per volume, then irregular particles are obtained. The second stage is performed

by placing the dispersion mixture in an aqueous solution of an inorganic salt, as previously described.

For a high concentration solution of co-monomers in the range of from about 50 percent to about 70 percent weight per weight, if a second dispersion stage is performed in alcohol, then spherical granules are obtained. Otherwise, if no second dispersion stage is performed, then irregular particles are obtained. For all of these structures, a similar washing procedure is performed as for the swollen block, except that an initial washing stage with glacial acetic acid is performed before the remaining washing steps.

After washing, the carboxylic cationites are preferably stored in water with 0.03% e-aminocaproic acid as a preservative, more preferably at a temperature of +4 °C in a refrigerator. For the testing of certain parameters and functional performance, the particulate cationite is dried in open air.

The preferred embodiment of the method of the present invention, in which low concentration co-monomer solution is employed, is optionally suitable for immobilization of a co-polymer inside another polymer matrix by simultaneous co-polymerization. According to this optional method, the first pre-polymer reaction mixture is prepared with a low concentration of co-monomers, preferably from a concentration of co-monomers of from about 20 percent to about 30 percent, weight per weight. This reaction mixture is then added to a medium containing the second monomer mixture, in which the medium contains alcohol with a long chain radical, such as butanol. Preferably, the first reaction mixture has a high viscosity, while the second monomer mixture has a lower viscosity. This optional but preferred method results in the formation of matrices of various balances between hydrophilic and hydrophobic properties, containing different functional groups, such as cationic and anionic groups in the co-polymers.

The properties of the cationites produced according to the method of the present invention result in improved sorption capacity for macromolecules such as proteins, and is suitable for isolation and purification of a wide range of proteins.

The ability of a carboxylic cationite to selectively and efficiently absorb macromolecules such as proteins is associated with the chemical nature of the employed co-monomers and the conditions for polymerization, which in turn affect the topology of the polymeric matrix. In order to increase the size of pores of the sorbent without substantially

reducing the volume concentration of the functional groups in the polymeric matrix, the copolymerization stage continues in a dispersion medium as previously described.

The components of the co-polymerization solution also have a significant effect on the topology and structure of the resultant polymeric matrix. For example, the intermolecular  
5 interaction of the elements of the structure leads to different topologies when an aqueous solution of acetic acid is used, in comparison with a mixture of acetic acid with an organic solvent. The polymer structures of the present invention are more hydrophobic than those disclosed in the background art. This increased hydrophobicity, and the correlation of the balance between hydrophilic and hydrophobic properties and the polymer structure, provide  
10 high sorption and improvement of chromatographic selectivity.

The carboxylic cationites prepared according to the method of the present invention are characterized by a number of functional parameters. For example, the swelling factor in H form in water and in a phosphate buffer by pH 7 was determined according to the following procedure. A sample of the swollen fraction of the cationites was placed into a graduated  
15 chromatographic column, and cationite volume was measured during the flow of water or phosphate buffer. The specific volume was determined to the weight of the cationites. The weight of sample was defined after drying the cationites, which swelled in water during the flow through the column. The loose weight of the cationite can be described as an index of porosity. The loose weight was measured according to the following procedure. A sample  
20 fraction of air-dried loose cationite with a defined fraction particle size of about 100 microns was placed in a cylinder, and the volume and weight of the sample was measured. The loose density was calculated by dividing the weight of the loose sample by the volume. The loose weight of the cationite of the present invention was found to preferably be in a range of from about 0.2 to about 0.6 g/ml or from about 0.06 to about 0.1 g/ml.

25 Exchange capacity with respect to sodium ions was defined by using 0.1 N sodium hydroxide in presence of 0.1 N sodium chloride solution.

The size of the pores was measured by scanning electron microscopy on prepared samples.

The carboxylic cationites prepared by abovementioned methods have insignificant  
30 swelling changes (typically a ratio of 1.2-1.8) due to transformation from the H<sup>+</sup> form into hydrogen-sodium form upon changing the pH value from 4 to 7, i.e. under the conditions of the sorption-desorption process. This property is in accordance with the requirement of

reversible sorption of proteins within the pH range from 4 to 7. The flow rate of a solution, containing proteins, which was passed through the column filled with a carboxylic cationite of the present invention, varied in the range of from about 200 to about 300 ml/hr/cm<sup>2</sup>, depending on the structure of the cationite, and if the cationite was in the H<sup>+</sup> form in the swollen state.

The carboxylic cationites of the present invention were found to be chemically stable in the pH range of from about 1 to about 14, such that they were stable through many cycles of regeneration, typically for about 100 cycles. The cationites were also found to be thermally stable at temperatures as high as 125 °C and as low as -20 °C. Carboxylic cationites prepared as particles and spherical granules have an exchange capacity with respect to sodium ions in a range of from about 20 meq/g to about 6 meq/g, and therefore from about 1.3 meq/ml to about 0.6 meq/ml.

The cationites of the present invention have advantages over the conventional ion exchange chromatography materials, CM Sephadex C25™ and CM Sephadex C50™, including higher sorption capacity for proteins, higher chemical stability, thermostability and greater ion exchange capacity. These parameters provide higher chromatographic selectivity for raw physiological liquids. Therefore, the cationites of the present invention are able to separate proteins of various natures and molecular weights from salt-containing solutions in a manner which is selective and fully reversible.

The carboxylic cationites prepared according to the present invention also have reproducible properties and superior sorption performances in comparison to the cationites prepared by the background art method. For example, the hemoglobin sorption capacity obtained with the carboxylic cationites of the present invention ranged from about 70 mg/ml (930 mg/g) to about 230 mg/ml (3830 mg/g) (Hb) while the sorption capacity achieved with the cationites prepared by dispersion in polysiloxane liquids was no more than 40 mg/ml (200-300 mg/g).

A further disadvantage of methods employing polysiloxane liquids is that the resulting cationite must be washed with petroleum, ether, acetones or other organic solvents. These solvents are flammable liquids and therefore appropriate precautions are required for safe performance of the background art process on an industrial scale. It can be readily appreciated that the additional required safety measures adds complications to the entire manufacturing process, thereby rendering the background art process more expensive and less economical.

Since the present invention preferably employs an aqueous dispersion medium and/or an alcohol dispersion medium, the resultant cationites do not require washing in flammable solvents, such that the manufacturing method is simple, cheap and easily adaptable to an industrial scale.

- 5           This invention will be further illustrated by the following Examples, which by no means limit the scope of the invention.

#### Example 1

- 10           A 150 ml flask was provided with a stirrer, an inlet pipe for argon and a dropping funnel. Into this flask, 10 ml of methacrylic acid, 1.22g of HTA and 45.5 ml of 70% acetic acid were poured. After dissolution of the co-monomers, argon was bubbled for 20 minutes. Then an initiator, 0.0569 g ammonium persulfate, was added. Copolymerization was conducted at a temperature of about 55-60 °C for 50 minutes. Next, the temperature was increased to 100 °C, and the mixture was held at that temperature for 1 hour. Next, after  
15           cooling to a temperature of 25 °C, the resultant swollen block of cationite was crushed, washed with water, a 1 N aqueous solution of sodium hydroxide at pH 8, a 1 N aqueous solution of hydrochloric acid and finally with water. The carboxylic cationite was then fractionated in the wet state.

- 20           The yield of cationite in the air-dry state was 10.9g (96%); the predominant size of particles was 100-200 microns. The exchange capacity with respect to sodium ion was found to be 10.23 meq/g of sorbent. Swelling factor of the cationite in the H-form in water was 4.6. The swelling factor of the cationite in the H-form in a phosphate buffer (pH 7) was 8.2. The specific volume of the cationite in the swollen state was 6.5 ml/g. The sorption capacity with respect to BSA (bovine serum albumin) (molecular mass of 66,000 Daltons) was 80 mg/ml,  
25           and was 75.4 mg/ml with respect to beta-globulin. The yield of protein upon desorption was found to be in a range of from about 97 % to about 100%. IgG<sub>1</sub> was isolated from raw mouse ascites liquid with this carboxylic cationite with a purity of 55.2% and a yield of 66%.

#### Example 2

- 30           A 150 ml flask was provided with a stirrer, an inlet pipe for argon and a dropping funnel. Into this flask, 10 ml of methacrylic acid, 133g of HTA and 45.5 ml of 93% acetic acid were poured. After dissolution of the co-monomers, argon was bubbled for 20 minutes.

Then an initiator, 0.0569 g ammonium persulfate, was added. Copolymerization was conducted at a temperature of about 70 °C for 40 minutes. Next, the temperature was increased to 100 °C, and the mixture was held at that temperature for 1 hour. Next, after cooling to a temperature of 25 °C, the resultant swollen block of cationite was crushed, washed with water, and then with a 1 M aqueous solution of hydrochloric acid. The carboxylic cationite was then fractionated in the wet state.

The yield of cationite in the air-dry state was 10.00 (89%); the predominant size of the particles was in the range of from about 100 to about 200 microns. Exchange capacity with respect to sodium ion was 9.87 meq/g of sorbent.

The swelling factor of the cationite in the H-form in water was 2 and in a phosphate buffer (pH 7) was 5.4. The specific volume of the swollen H-form in water was 9.21 ml/g and in phosphate buffer (pH 7) was 24.8 ml/g.

IgG<sub>1</sub> was isolated from raw mouse ascites liquid with this carboxylic cationite in a one-step process with a purity of 88.7% and a yield of 78%.

### Example 3

A 150 ml flask was provided with a stirrer, an inlet pipe for argon and a dropping funnel. Into this flask, 10 ml of methacrylic acid, 1.22g of HTA and 45.5 ml of 80% acetic acid were poured. After dissolution of the co-monomers, argon was bubbled for 20 minutes. Then an initiator, 0.0569 g ammonium persulfate, was added. Co-polymerization was conducted at a temperature of from about 55 to about 60 °C for 50 minutes. Next, the temperature was increased to 100 °C, and the mixture was held at that temperature for 1 hour. Next, after cooling to a temperature of 25 °C, the resultant swollen block of cationite was crushed, washed with water, a 1 N aqueous solution of sodium hydroxide at pH 8, a 1 N aqueous solution of hydrochloric acid and finally with water. The carboxylic cationite was then fractionated in the wet state.

The yield of cationite in the air-dry state was 10.1g (90%); the predominant size of particles was in a range of from about 100 to about 200 microns. The exchange capacity with respect to sodium ion was 10.00 meq/g of sorbent. The swelling factor of the cationite in the H-form in water was 1.2 and in a phosphate buffer (pH 7) was 2. The specific volume of the swollen H-form in water was 6.1 ml/g and in phosphate buffer (pH 7) was 10.1 ml/g.

IgG<sub>1</sub> was isolated from raw mouse ascites liquid with this carboxylic cationite in a one-step process with a purity of 57.6% and a yield of 78%.

#### Example 4

5 A 150 ml flask was provided with a stirrer, an inlet pipe for argon and a dropping funnel. Into this flask, 10 ml of methacrylic acid, 1.22g of HTA and 45.5 ml of a solution of water and 1 ml of polyethyleneglycol were poured. After dissolution of the co-monomers, argon was bubbled for 20 minutes. Then an initiator, 0.056 g ammonium persulfate, was added. Copolymerization was conducted at a temperature of from about 50 to about 60 °C for 10 50 minutes. Next, the temperature was increased to 100 °C, and the mixture was held at that temperature for 1 hour. Next, after cooling to a temperature of 25 °C, the resultant swollen block of cationite was crushed, washed with water, a 1 N aqueous solution of sodium hydroxide at pH 8, a 1 N aqueous solution of hydrochloric acid and finally with water. The carboxylic cationite was then fractionated in the wet state.

15 The yield of cationite in the air-dry state was 10.9 g (96%); the predominant size of particles was in a range of from about 100 to about 200 microns. The exchange capacity with respect to sodium ion was 10.01 meq/g of sorbent. The swelling factor of the cationite in the H-form in water was 2.8 and in a phosphate buffer (pH 7) was 4. The specific volume of the swollen H-form in water was 4.3 ml/g and in phosphate buffer (pH 7) was 6.2 ml/g.

20 IgG<sub>1</sub> was isolated from desalted mouse ascites liquid with this carboxylic cationite in a one-step process with a purity of 40 % and a yield of 98.3 %. The sorption capacity of IgG<sub>1</sub> contaminating proteins for isolation from raw ascites fluid was about 40%, such that this cationite may be used for a preliminary step of purification of IgG<sub>1</sub>.

#### Example 5

25 A 250 ml flask was provided with a stirrer, an inlet pipe for argon and a dropping funnel. Into this flask, 180 ml of 20% sodium sulfate was poured and argon was bubbled for 30 minutes. At the same time, 10 ml of methacrylic acid, 1.22g of HTA and 45.5 ml of 70% acetic acid was charged into the funnel in a current of argon. After dissolution of the co- 30 monomers, an initiator, 0.0569 g ammonium persulfate, was added. Copolymerization was conducted at a temperature of about 60 °C for 8 minutes to obtain a high viscosity pre-polymer. The pre-polymer was dispersed with 20 ml of butanol as the dispersion medium.



After stirring, the resultant dispersion was poured into 180 ml of a solution of 20% sodium persulfate. The temperature was maintained at 60 °C for 30 minutes. Next, the temperature was increased to 100 °C, and the mixture was held at that temperature for 1 hour.

Next, after cooling to a temperature of 25 °C, the resultant particles of cationite were separated from the dispersion phase. The particles were then washed with glacial acetic acid, then with water, next with a 1 N aqueous solution of sodium hydroxide at pH 8, then with a 1 N aqueous solution of hydrochloric acid and finally with water. The carboxylic cationite was then fractionated in the wet state.

The predominant particle size was 100 microns in the globular form, and otherwise 200 x 100 microns. The exchange capacity with respect to sodium ion was 1.3 meq/ml, 21.3 meq/g of sorbent. The specific volume of the swollen H-form in water was 16 ml/g and in phosphate buffer (pH 7) was 22.4 ml/g.

The sorption capacity of this cationite with respect to Hb (hemoglobin, molecular weight of 66,000 Daltons) was 230 mg/ml, 3830 mg/g of sorbent.

#### Example 6

A 250 ml flask was provided with a stirrer, an inlet pipe for argon and a dropping funnel. Into this flask, 140 ml of 20% sodium sulfate was poured and argon was bubbled for 30 minutes. At the same time, 5 ml of methacrylic acid, 0.61 g of HTA and 5 ml of 5% acetic acid was charged into the funnel in a current of argon. After dissolution of the co-monomers, a mixture of 0.044 g ammonium persulfate and 0.015 sodium methabisulfite was added as the initiator. Copolymerization was conducted at a temperature of about 22 °C for 8 minutes to obtain a pre-polymer. The pre-polymer was dispersed with 50 ml of an aqueous solution of 20% sodium sulfate as the dispersion medium. While stirring, the temperature was maintained at 22 °C for 20 minutes. The temperature was increased to 50 °C for 10 minutes, and then to 60 °C for 20 minutes. Next, the temperature was increased to 100 °C, and the mixture was held at that temperature for 1 hour.

Next, after cooling to a temperature of 25 °C, irregular particles of cationite were separated from the dispersion phase by a water-jet pump. The particles were then washed with glacial acetic acid, then with water, next with a 1 N aqueous solution of sodium hydroxide at pH 8, then with a 1 N aqueous solution of hydrochloric acid and finally with water. The carboxylic cationite was then fractionated in the wet state.

The predominant particle size was 200 x 100 microns. The exchange capacity with respect to sodium ion was 1.2 meq/ml or 12.8 meq/g. The specific volume of the swollen H-form in water was 10.5 ml/g and in phosphate buffer (pH 7) was 13.5 ml/g.

#### 5 Example 7

A 250 ml flask was provided with a stirrer, an inlet pipe for argon and a dropping funnel. Into this flask, 140 ml of an aqueous solution of 20% sodium sulfate was poured and argon was bubbled for 30 minutes. At the same time, 5 ml of methacrylic acid, 0.61 g of HTA and 5 ml of 70% acetic acid was charged into the funnel in a current of argon. After  
10 dissolution of the co-monomers, 0.0569 g ammonium persulfate was added as the initiator. Co-polymerization was conducted at a temperature of about 55 °C for 15 minutes to obtain a pre-polymer. The pre-polymer was dispersed with 50 ml of an aqueous solution of 20% sodium sulfate as the dispersion medium. The mixture was stirred for 5 minutes without heating. The temperature was then increased to 70 °C for 45 minutes, and then to 90 °C for  
15 30 minutes. 17 ml of butanol was then added, and the temperature was maintained for 30 minutes. Next, the temperature was increased to 100 °C, and the mixture was held at that temperature for 30 minutes.

Next, after cooling to a temperature of 25 °C, granules of cationite were separated from the dispersion phase by a water-jet pump. The particles were then washed with glacial  
20 acetic acid, then with water, next with a 1 N aqueous solution of sodium hydroxide at pH 8, then with a 1 N aqueous solution of hydrochloric acid and finally with water. The carboxylic cationite was then fractionated in the wet state.

The predominant particle size was in a range of from about 20 to about 70 microns. The exchange capacity with respect to sodium ion was 0.83 meq/ml or 11.2 meq/g. The  
25 specific volume of the swollen H-form in water was 12.5 ml/g and in phosphate buffer (pH 7) was 23 ml/g.

#### Example 8

A 250 ml flask was provided with a stirrer, an inlet pipe for argon and a dropping  
30 funnel. Into this flask, 140 ml of 20% sodium sulfate was poured and argon was bubbled for 30 minutes. At the same time, 5 ml of methacrylic acid, 0.61 g of HTA and 22.8 ml of 5% acetic acid were charged into the funnel in a current of argon. After dissolution of the co-

monomers, a mixture of 0.044 g ammonium persulfate and 0.015 sodium methabisulfite was added as the initiator. Copolymerization was conducted at a temperature of about 22 °C for 15 minutes to obtain a pre-polymer.

5 The prepolymer was dispersed with 23 ml of an organic phase, which included 20.2 ml (16.34 g) of butanol and 2.8 ml (2.3 g) of lauric alcohol. After stirring, the dispersion was poured into 140 ml of a 20% sodium sulfate aqueous solution.

Stirring was continued for 5 minutes, then the process was allowed to incubate at 22 °C for 5 hours. The flask temperature was then increased to 48-55 °C and was held at 55 °C for an hour. Stirring was then stopped overnight. The heating process with stirring was  
10 resumed the next day, starting at a temperature of 50 °C, with an increase in temperature of 10 °C every 0.5 hour to a final temperature of 100 °C, which was maintained for 30 minutes.

Next, after cooling to a temperature of 25 °C, granules of cationite were separated from the dispersion phase. The granules were then washed with glacial acetic acid, then with water, next with a 1 N aqueous solution of sodium hydroxide at pH 8, then with a 1 N  
15 aqueous solution of hydrochloric acid and finally with water.

The total binding capacity of hemoglobin was 74 mg/ml or 908 mg/g. The specific volume of the swollen H-form in water was 12.3 ml/g and in phosphate buffer (pH 7) was 18.4 ml/g.

## 20 Example 9

A 250 ml flask was provided with a stirrer, an inlet pipe for argon and a dropping funnel. Into this flask, 140 ml of 20% sodium sulfate was poured and argon was bubbled for 30 minutes. At the same time, 5 ml of methacrylic acid, 0.61 g of HTA and 22.8 ml of 70% acetic acid were charged into the funnel in a current of argon. After dissolution of the co-  
25 monomers, 0.0569 g ammonium persulfate was added as the initiator. Copolymerization was conducted at a temperature of about 60 °C for 50-60 minutes to obtain a pre-polymer.

The prepolymer was dispersed with 23 ml of an organic phase, which included 20.2 ml (16.34 g) of butanol, 2.8 ml (2.3 g) of lauric alcohol and 0.23 g oleic acid. After stirring, the dispersion was poured into 140 ml of a 20% sodium sulfate aqueous solution.

30 Stirring was continued for 5 minutes, and then the flask temperature was increased to 60 °C for 30 minutes. The flask temperature was then increase to 70 °C over 30 minutes, with

an increase in temperature of 10 °C every 40 minutes to a final temperature of 100 °C, which was maintained for 30 minutes.

Next, after cooling to a temperature of 25 °C, granules of cationite were separated from the dispersion phase. The granules were then washed with glacial acetic acid, then with water, next with a 1 N aqueous solution of sodium hydroxide at pH 8, then with a 1 N aqueous solution of hydrochloric acid and finally with water. The carboxylic cationite was then fractionated in the wet state.

The predominant particle size was in a range of from about 40 to about 120 microns. The exchange capacity with respect to sodium ion was 0.56 mg/ml. The specific volume of the swollen H-form in water was 30 ml/g and in phosphate buffer (pH 7) was 60 ml/g. The total binding capacity of hemoglobin was 94.7 mg/ml or 2841 mg/g.

#### Example 10

The chemical and physical properties of two of these carboxylic cationites of the present invention, #27 and #44, were examined in further detail. Cationites #27 and #44 were prepared as swollen blocks. Cationite #27 was prepared as for Example 1, while cationite #44 was prepared as for Example 4.

First, water regain was examined for sorbents in the H<sup>+</sup> form in sodium phosphate buffer as a function of the pH. Water regain was found to reach a steady state at pH 6 for #44 and at pH 7 for #27. The results are summarized in Table 1.

Table 1. Water regain as a function of pH

| <u>Buffer &amp; pH</u> | <u>#27</u> | <u>#44</u> |
|------------------------|------------|------------|
| phosphate buffer, pH 3 | 100        | 100        |
| phosphate buffer pH 6  | 150        | 125        |
| phosphate buffer pH 7  | 185        | 125        |
| phosphate buffer pH 8  | 185        | 125        |

The mechanical stability and elasticity of these cationites was then tested as follows.

Bed volume was measured in correlation with flow rate, which was sustained by a peristaltic (0.28 cm<sup>2</sup> x 10 cm column). The reduction in volume was about 15% to about 20% of the initial volume, up to a flow rate of about 350 ml/h<sup>-1</sup>/cm<sup>-2</sup>.

The chemical stability of cationites #27 and #44 against oxidative agents was then tested by incubating these cationites with 0.3 g/l NaOCl (RT) for about 16 hours. The water regain and H<sup>+</sup> ion binding capacity were then examined. Both were found to have remained substantially unchanged (maximum difference of 6% and 5% respectively).

- 5        These cationites were also tested for their ability to undergo regeneration cycles, for example with 0.5-1.0 M NaOH and 0.1 M HCl, ethanol or various organic solvents. The average regeneration cycle number while maintaining sorbing capacity was found to be about 100.

10    Example 11

The chromatographic resolving properties of two of the carboxylic cationites of the present invention, #27 and #44, were examined in further detail, in comparison to CM Sepharose™ (Pharmacia Ltd., hereinafter referred to as "commercial product #1") and Bio-Rex 70™ (Bio-Rad Ltd., 100-200 mesh, hereinafter referred to as "commercial product #2").

- 15    The experimental method was as follows.

Three fold diluted rabbit serum proteins were loaded on columns packed with #27, #44, commercial product #1 or commercial product #2.

- 20    For #27 or #44, cationites of the present invention, adsorption was performed in a buffer containing 10 mM sodium acetate, pH 5. Desorption was performed in a salt gradient of 0-0.75 M ammonium acetate in 20 mM Tris, pH 7.2. Fractions numbered 3-8 were collected as shown by the arrows in Figure 1A, and then loaded on an SDS-PAGE gel, shown in Figure 1B. The lanes of the SDS-PAGE gel were loaded as follows: lane 1 - molecular weight markers; lane 2 - serum polypeptides before separation; lanes 3-8 correspond to the collected fractions. Figures 1A and 1B are shown for cationite #27, but a similar separation  
25    profile was obtained for cationite #44.

As can be seen in Figures 1A and 1B, fractions 8-10 contain the albumin fraction, while fractions 10-24 contain IgG<sub>1</sub>. None of the protein eluted before the salt gradient.

- 30    For commercial product #1, adsorption was performed in 10 mM sodium acetate, pH 4.8 or 20 mM phosphate buffer, pH 7.2. Desorption was performed in a continuous salt gradient of 0-0.5 M NaCl in 20 mM phosphate buffer pH 7.2. Fractions were collected as shown by the arrows in Figure 2A, and were then loaded on an SDS-PAGE gel as shown in Figure 2B. Lanes are as for Figure 1B. Essentially, most of the protein eluted before the salt

gradient and did not bind to the resin. Some separation of serum proteins occurred at pH 4.8, but none occurred at pH 7.2.

For commercial product #2, adsorption was performed in 10 mM phosphate buffer, pH 6.5. Desorption was performed in a continuous salt gradient of 0-0.75 M NaCl in 10 mM phosphate buffer pH 6.5. Fractions were collected as shown in Figure 3A, and were then loaded on an SDS-PAGE gel as shown in Figure 3B. Lanes are as for Figure 1B, except that the fraction number is indicated below each lane. Essentially, most of the protein eluted before the salt gradient and did not bind to the resin. Furthermore, the majority of both albumin and IgG<sub>1</sub> eluted in a single peak, which is the first peak as shown.

### Example 12

Protein purity and recovery was tested by using several carboxylic cationite sorbents of the present invention, commercial product #1, DEAE cellulose DE-52 (hereinafter referred to as "commercial product #3") and CM Sephadex C-25 and C-50<sup>TM</sup> (Pharmacia Ltd., hereinafter referred to as "commercial product #4" and "commercial product #5", respectively).

The first test was performed to recover Immunoglobulin G<sub>1</sub> (Ig G<sub>1</sub>) from initial ascites mouse fluid diluted with 2 M ammonium sulfate, pH 6.32, 1:1 volume per volume. The column was 0.78 x 4.2 cm, packed with 2 ml of the sorbent. The flow rate was 37 ml/h/cm<sup>2</sup>. The sorbent was regenerated by using 1 M NaOH, followed by 1 M HCl, followed by water, followed by equilibration with 0.1 M acetate buffer, pH 5. Desorption was performed with a stepwise pH gradient of 0.1 M citrate buffer, pH 5 to 7.7, to pH 7.7 buffer with 0.2 M NaCl.

No purification was observed with commercial product #2. Carboxylic cationites #22 and #28 of the present invention were tested. Results are shown in Table 2. Cationite #22 was prepared as for Example 2, while cationite #28 was prepared as for Example 3.

Table 2

| Sorbent | immunoglobulin<br>in peaks (μg) | recovery<br>(peak %) | purity, % | desorption<br>(total μg) | total<br>recovery, % |
|---------|---------------------------------|----------------------|-----------|--------------------------|----------------------|
| #22     | 89                              | 5.6                  | 2.6       | 484                      | 30.7                 |
|         | 395                             | 25                   | 9.3       |                          |                      |

|     |     |      |      |     |      |
|-----|-----|------|------|-----|------|
| #22 | 57  | 3.6  | 9.7  | 446 | 28.2 |
|     | 29  | 1.8  | 0.5  |     |      |
|     | 354 | 22.4 | 88.7 |     |      |
| #28 | 25  | 1.6  | 1.5  | 730 | 46.2 |
|     | 136 | 8.6  | 4.9  |     |      |
|     | 567 | 35.9 | 57.6 |     |      |

The next test was performed to recover Immunoglobulin G<sub>1</sub> (Ig G<sub>1</sub>) from desalted ascites mouse fluid diluted with 0.1 M acetic acid, pH 5, 1:1 volume per volume. The column was 0.78 x 3.8 cm, packed with 2 ml of the sorbent. The flow rate was 37 ml/h/cm<sup>2</sup>. The sorbent was regenerated by using 1 M NaOH, followed by 1 M HCl, followed by water, followed by equilibration with 0.1 M acetate buffer, pH 5. Desorption was performed with a stepwise pH gradient of 0.1 M citrate buffer, pH 5 to 6.

Commercial product #3 and carboxylic cationites #26, #27, #36, #37, and #44 of the present invention were tested. Results are shown in Table 3.

**Table 3**

| <u>Sorbent</u>            | <u>immunoglobulin</u><br><u>in peaks (μg)</u> | <u>recovery</u><br><u>(peak %)</u> | <u>purity, %</u> | <u>desorption</u><br><u>(total μg)</u> | <u>total</u><br><u>recovery, %</u> |
|---------------------------|---|------------------------------------|------------------|--|------------------------------------|
| #26                       | 1,220   | 54.1                               | 69.2             | 1,705                                  | 62.0                               |
|                           | 0.267   |                                    | 100              |  |                                    |
| #27                       | 2,290   | 83.3                               | 66.0             | 2,368                                  | 86.1                               |
| #36                       | 2,317   | 95.9                               | 62.3             | 2,782                                  | 101.2                              |
|                           | 0.319   |                                    | 62.2             |  |                                    |
| #37                       | 1,337   | 72.5                               | 100              | 2,092                                  | 76.1                               |
|                           | 0.656   |                                    | 16.8             |  |                                    |
| #44                       | 2,419   | 88.0                               | 39.5             | 2,701                                  | 98.3                               |
| commercial<br>product # 3 | 2,613   | 95.1                               | 30.1             | 2,749                                  | 100                                |

Table 4 shows a comparison between the efficacy of carboxylic cationites #25, #26, #27, #28, #30, #36 and 37 of the present invention for either desalted or initial mouse ascites fluid. Cationite #25 was prepared with 20 percent weight per weight co-monomers, which were methacrylic acid and HTA as the cross-linking agent, with 1,1' azobis (cyclohexanecarbonitrile) as the initiator, in a solution of glacial acetic acid (100%) to form a swollen block. Cationite #27 was prepared with 5 percent acetic acid in octyl alcohol. Cationite #27 was prepared with 70 percent volume per volume acetic acid in water solution. Cationite #30 was prepared as for cationite #25, except that the co-polymerization solution was 93% acetic acid and 7% dimethylformamide, volume per volume. Cationite #36 was prepared from 20 percent dimethylformamide volume per volume in acetic acid. Cationite #37 was prepared with 5 percent volume per volume acetic acid in butyl alcohol.

Max purity refers to the purity % in the purest peak. Max recovery refers to the percentage recovery in purest peak alone. and Desor. ( $\mu\text{g}$ ) refers to the total protein desorption in micrograms.

Table 4

| <u>sorbent</u> | <u>desalted ascites</u>     |                               |  | <u>initial ascites</u>      |                               |  |
|----------------|-----------------------------|-------------------------------|--|-----------------------------|-------------------------------|--|
|                | <u>max</u><br><u>purity</u> | <u>max</u><br><u>recovery</u> | <u>desor. (<math>\mu\text{g}</math>)</u> | <u>max</u><br><u>purity</u> | <u>max</u><br><u>recovery</u> | <u>desor. (<math>\mu\text{g}</math>)</u> |
| #25            | 4.7                         | 15.3                          | 241                                      | 11.7                        | 3.8                           | 208                                      |
| #27            | 33.5                        | 58.3                          | 1,247                                    | 6.5                         | 10.3                          | 339                                      |
| #28            | 100                         | 16                            | 499                                      | 57.6                        | 35.9                          | 730                                      |
| #30            | 17.4                        | 40.2                          | 636                                      | 7.4                         | 22.4                          | 354                                      |

The next test was performed in a two-step purification to recover Immunoglobulin G<sub>1</sub> (Ig G<sub>1</sub>) from desalted ascites mouse fluid diluted with water, pH 7.25, 1:1 volume per volume. The column was 0.78 x 3.8 cm, packed with 1 ml of the sorbent. The flow rate was 37 ml/h/cm<sup>2</sup>. The sorbent was regenerated by using 1 M NaOH, followed by 1 M HCl, followed by water, followed by equilibration with 0.1 M acetate buffer, pH 5. Desorption was



performed with a stepwise pH gradient of 0.1 M citrate buffer, pH 5 to 8.2. In the second step, unsorted eluted fractions 1-12 from a column of cationite #44 were loaded onto a column with cationite #27. The comparison was to commercial product #1. Results are shown in Table 5.

5

Table 5

| <u>Sorbent</u>            | <u>immunoglobulin</u><br><u>in peaks (μg)</u> | <u>recovery</u><br><u>(peak %)</u> | <u>purity, %</u> | <u>desorption</u><br><u>(total μg)</u> | <u>total</u><br><u>recovery, %</u> |
|---------------------------|---|------------------------------------|------------------|--|------------------------------------|
| First step -<br>#44       | 19.1  | 0.3                                | 0.7              | 19.1                                   | 0.7                                |
| Second step -<br>#27      | 5,275   | 86.3                               | 75.4             | 5,310                                  | 86.6                               |
| commercial<br>product # 1 | 4,253   | 69.5                               | 65.1             | 4,300                                  | 70.3                               |

The next test was performed in a two-step purification to recover Immunoglobulin G<sub>1</sub> (Ig G<sub>1</sub>) from dialyzed ascites mouse fluid diluted with water, pH 7.25, 1:1 volume per volume. The column was 0.78 x 3.8 cm, packed with 1 ml of the sorbent. The flow rate was 37 ml/h/cm<sup>2</sup>. The sorbent was regenerated by using 1 M NaOH, followed by 1 M HCl, followed by water, followed by equilibration with 0.1 M acetate buffer, pH 5. Desorption was performed with a stepwise pH gradient of 0.1 M citrate buffer, pH 5 to 8.2. In the second step, unsorted eluted fractions 1-5 from a column of cationite #44 were loaded onto a column of cationite #27. The comparison was to commercial product #1. Results are shown in Table

10

15 6.

Table 6

| <u>Sorbent</u>      | <u>immunoglobulin</u><br><u>in peaks (μg)</u> | <u>recovery</u><br><u>(peak %)</u> | <u>purity, %</u> | <u>desorption</u><br><u>(total μg)</u> | <u>total</u><br><u>recovery, %</u> |
|---------------------|---|------------------------------------|------------------|--|------------------------------------|
| First step -<br>#44 | 5,829   | 63.0                               | 100              | 8,617                                  | 93.2                               |
|                     | 2,789   | 30.2                               | 42.5             |  |                                    |

|               |       |      |      |       |      |
|---------------|-------|------|------|-------|------|
| Second step - | 267   | 2.9  | 44.1 | 267   | 2.9  |
| #27           |       |      |      |       |      |
| commercial    | 4,362 | 47.2 | 59.9 | 6,780 | 73.3 |
| product # 1   |       |      |      |       |      |
|               | 2,418 | 26.1 | 35.5 |       |      |

The next test was performed to recover Immunoglobulin G<sub>2A</sub> (Ig G<sub>2A</sub>) from initial ascites mouse fluid diluted with water, pH 8.256, 2:1 volume per volume, electroconductivity 5.5 except for commercial product #1-2 (CM Sepharose Fast Flow - 2), where the dilution ratio was 6:1 volume per volume, electroconductivity 2.8. The column was 0.78 x 3.8 cm, packed with 1 ml of the sorbent. The flow rate was 37 ml/h/cm<sup>2</sup>. The cationite was regenerated by using 1 M NaOH, followed by 1 M HCl, followed by water, followed by equilibration with 0.1 M acetate buffer, pH 5. Desorption was performed with a stepwise pH gradient of 0.1 M citrate buffer, pH 5 to 8.2.

Commercial product #1 and cationites #27 and #44 were tested. Results are shown in Table 7.

Table 7

| <u>Sorbent</u> | <u>immunoglobulin</u><br><u>in peaks (μg)</u> | <u>purity, %</u> | <u>desorption</u><br><u>(total μg)</u> | <u>total</u><br><u>recovery,</u><br><u>%</u> |
|----------------|---|------------------|--|--|
| #27            | 1,272   | 10.1             | 1,377                                  | 49.2   |
|                | 51  | 1.8              |  |  |
|                | 53  | 1.9              |  |  |
| #44            | 1,425   | 10.5             | 1,514                                  | 54.1   |
|                | 90  | 2.1              |  |  |
| commercial     | 484   | 5.5              | 537                                    | 19.2   |
| product #1-1   | 53  | 27.9             |  |  |
| commercial     | 1,006   | 5.5              | 1,006                                  | 35.9   |
| product # 1-2  |   |                  |  |  |

The next test was performed to separate a mixture of human and goat serum albumin diluted with acetic buffer, pH 5.8, 1:1 weight per weight. The column was 0.02 x 0.9 cm, packed with 1 ml of the cationite sorbent. The flow rate was 37 ml/h/cm<sup>2</sup>. The sorbent was regenerated by using 1 M NaOH, followed by 1 M HCl, followed by water, followed by equilibration with 0.1 M acetate buffer, pH 5. Desorption was performed with a stepwise pH gradient of 0.5 M ammonium acetate buffer, pH 4.5 to 7. Carboxylic cationite #27 was tested. Results are shown in Table 8.

Table 8

| <u>Sorbent</u> | <u>Protein solution:</u><br><u>sorbent (v:v)</u> | <u>protein pH</u> | <u>purity of</u><br><u>main peak</u><br><u>%</u> | <u>recovery, %</u> | <u>minimum #</u><br><u>of peaks</u> |
|----------------|--|-------------------|--|--------------------|-------------------------------------|
| #27 (1)        | 4.6:1  | 4.7               | 78   | 96                 | 5                                   |
| #27 (2)        | 18:1   | 5.8               | 89   | 20                 | 1                                   |

Next, a graph was prepared for such a separation of a mixture of human (HSA) and goat (GSA) serum albumin diluted with acetic buffer, pH 4.7, 1:1 weight per weight, using cationite #27, shown in Figure 4. The column was 0.02 cm<sup>2</sup> x 3 cm, packed with 5 ml of the cationite sorbent. The flow rate was 37 ml/h/cm<sup>2</sup>. The sorbent was regenerated by using 1 M NaOH, followed by 1 M HCl, followed by water, followed by equilibration with 0.1 M acetate buffer, pH 4.7. Desorption was performed with a stepwise pH gradient of 0.5 M ammonium acetate buffer, pH 5 to 7.5. Cationite #27 was tested.

Figure 5 shows a graph of a separation of non-treated fresh milk proteins, in which the calcium concentration was 1.25 mg/ml. The column was 2 cm<sup>2</sup> x 0.6 cm, packed with 1 ml of the cationite sorbent. The flow rate was 37 ml/h/cm<sup>2</sup>. The sorbent was regenerated by using 1 M NaOH, followed by 1 M HCl, followed by water, followed by equilibration with 0.1 M acetate buffer, pH 4.7. Desorption was performed with a stepwise pH gradient of 0.5 M ammonium acetate buffer, pH 5 to 7.7. Cationite #27 was tested.

Figure 6 shows a graph of a separation of human blood plasma proteins, diluted with water 2:1 (weight per weight). The column was 2 cm<sup>2</sup> x 0.6 cm, packed with 1 ml of the cationite sorbent. The flow rate was 37 ml/h/cm<sup>2</sup>. The sorbent was regenerated by using 1 M NaOH, followed by 1 M HCl, followed by water, followed by equilibration with 0.1 M

acetate buffer, pH 4.7. Desorption was performed with stepwise pH gradients of pH (7.7-10.8), ammonium acetate concentration (0.25 to 0.3 M), combined with ammonium chloride and ammonium hydroxide. Carboxylic cationite #27 of the present invention was tested.

5. It will be appreciated that the above descriptions are intended only to serve as examples, and that many other embodiments are possible within the spirit and the scope of the present invention.

## WHAT IS CLAIMED IS:

1. A method for producing a carboxyl cationite, comprising the steps of:
  - (a) dissolving a monovinyl monomer and a cross-linking agent as co-monomers in a first solvent to form a first mixture, said first solvent being selected from the group consisting of a mixture of an organic solvent and water, a mixture of an organic solvent and acetic acid, and an aqueous solution of acetic acid wherein an amount of said acetic acid is in a range of from about 60% to about 100%, volume per volume; and
  - (b) adding an initiator of radical polymerization to said first mixture to form a swollen block of the carboxyl cationite, thereby obviating the need for dispersing the carboxyl cationite in a dispersing medium.
2. The method of claim 1, wherein said first solvent is selected from the group consisting of a mixture of an organic solvent and water, and a mixture of an organic solvent and acetic acid, and said organic solvent is selected from the group consisting of dimethylformamide, monobutyl glycolate, polyethylene glycol and an alcohol.
3. The method of claim 2, wherein said alcohol is selected from the group consisting of butyl alcohol, octyl alcohol and decyl alcohol.
4. The method of claim 2, wherein said first solvent is said mixture of said organic solvent and said acetic acid, and wherein said acetic acid is present in an amount of from about 20% to about 50% volume per volume.
5. The method of claim 1, wherein said first solvent is said aqueous solution of acetic acid, wherein said amount of said acetic acid is in said range of from about 60% to about 100%, volume per volume.
6. The method of claim 5, wherein said monovinyl monomer is selected from the group consisting of a methacrylic acid, an acrylic acid and a methacryloylaminocarbonic acid.

7. The method of claim 6, wherein said monovinyl monomer is selected from the group consisting of methacrylic acid, acrylic acid, methacryloylcaproic acid, methacryloylaminobenzoic acid, methacryloylglycine and methacryloylaminocarbonic acid.

8. The method of claim 7, wherein an amount of said monovinyl monomer and an amount of said cross-linking agent are together in a range of from about 10 percent to about 30 percent weight per weight of said first mixture.

9. The method of claim 8, wherein said cross-linking agent is a long chain cross-linking agent.

10. The method of claim 9, wherein said long chain cross-linking agent is selected from the group consisting of hexahydro-1,3,5-triacryloyl triazine (HTA), N,N'-methylene diacrylamide (MDAA), N,N'-ethylene dimethacrylamide (EDMA), N,N'-hexamethylene dimethacrylamide (HMDMA), triallyl isocyanurate (TAIC), *p*-phenylene dimethacrylamide (*p*-PHDMA), and benzidine dimethacrylamide (BDMA).

11. The method of claim 10, wherein said long chain cross-linking agent is selected from the group consisting of HTA, EDMA, and TAIC.

12. The method of claim 11, wherein said initiator is selected from the group consisting of ammonium persulfate, a 1,1' azobis (cyclohexanecarbonitrile), and a combination of ammonium persulfate - sodium methabisulfite.

13. The method of claim 12, wherein step (b) is performed at a temperature in a range of from about 20 to about 25 °C if said initiator is said combination of ammonium persulfate - sodium methabisulfite, or alternatively at a temperature in a range of from about 50 to about 60 °C if said initiator is ammonium persulfate or a 1,1' azobis (cyclohexanecarbonitrile).

14. The method of claim 1, further comprising the steps of:

- (c) crushing said swollen block of the carboxylic cationite to form particulate matter; and
- (d) washing said particulate matter.

15. The method of claim 14, wherein step (d) includes the steps of:

- (i) washing said particulate matter with water;
- (ii) washing said particulate matter with an aqueous solution of from about 0.5 N to about 1 N sodium hydroxide of pH 8;
- (iii) washing said particulate matter with an aqueous solution of about 1 N hydrochloric acid; and
- (iv) washing said particulate matter with water.

16. A method for forming particulate matter of carboxylic cationites, the method comprising the steps of:

- (a) dissolving a monovinyl monomer and a cross-linking agent as co-monomers in a first solvent to form a first mixture;
- (b) adding an initiator of radical polymerization to said first mixture to form a pre-polymer;
- (c) dispersing said pre-polymer in a solution of alcohol to form a dispersion mixture; and
- (d) mixing said dispersion mixture with an aqueous solution of an inorganic salt to form the particulate matter.

17. The method of claim 16, wherein an amount of said monovinyl monomer and an amount of said cross-linking agent are together in a range of from about 20 percent to about 30 percent weight per weight of said first mixture.

18. The method of claim 17, wherein said first solvent is selected from the group consisting of an aqueous solution of acetic acid and a mixture of an organic solvent with acetic acid.

19. The method of claim 18, wherein said first solvent is said aqueous solution of acetic acid, and said aqueous solution of acetic acid includes an amount of acetic acid in a range of from about 5 percent to about 15 percent volume per volume.

20. The method of claim 18, wherein said first solvent is said aqueous solution of acetic acid, and said aqueous solution of acetic acid includes an amount of acetic acid in a range of from about 60 percent to about 100 percent volume per volume.

21. The method of claim 18, wherein said first solvent is said mixture of said organic solvent and said acetic acid, and said acetic acid is present in an amount of from about 20% to about 50% volume per volume.

22. The method of claim 21, wherein said organic solvent is selected from the group consisting of dimethylformamide, monobutyl glycolate and polyethylene glycol.

23. The method of claim 22, wherein said alcohol is selected from the group consisting of butyl alcohol, octyl alcohol and decyl alcohol.

24. The method of claim 23, wherein a ratio of said pre-polymer to said alcohol is in a range of from about 2.5:1 to about 5:1.

25. The method of claim 23, wherein a ratio of said pre-polymer to said alcohol is about 1:1.

26. The method of claim 23, wherein a ratio of said dispersion mixture to said aqueous solution of said inorganic salt is in a range of from about 1:4 to about 1:5.

27. The method of claim 26, wherein said aqueous solution of said inorganic salt is an aqueous solution of sodium sulfate.

28. The method of claim 27, wherein an amount of said sodium sulfate in said aqueous solution of sodium sulfate is about 20 percent weight per weight.



29. The method of claim 16, wherein said monovinyl monomer is selected from the group consisting of a methacrylic acid, an acrylic acid and a methacryloylaminocarbonic acid.

30. The method of claim 29, wherein said monovinyl monomer is selected from the group consisting of methacrylic acid, acrylic acid, methacryloylcaproic acid, methacryloylaminobenzoic acid, methacryloylglycine and methacryloylaminocarbonic acid.

31. The method of claim 30, wherein said cross-linking agent is a long chain cross-linking agent.

32. The method of claim 31, wherein said long chain cross-linking agent is selected from the group consisting of hexahydro-1,3,5 -triacyloyl triazine (HTA), N,N'-methylenediacrylamide (MDAA), N,N'-ethylenedimethacrylamide (EDMA), N,N'-hexamethylenedimethacrylamide (HMDMA), triallylisocyanurate (TAIC), *p*-phenylenedimethacrylamide (*p*-PHDMA), and benzidinedimethacrylamide (BDMA).

33. The method of claim 32, wherein said long chain cross-linking agent is selected from the group consisting of HTA, EDMA, and TAIC.

34. The method of claim 33, wherein said initiator is selected from the group consisting of ammonium persulfate, a 1,1' azobis (cyclohexanecarbonitrile), and a combination of ammonium persulfate - sodium methabisulfite.

35. A method for forming particulate matter of a carboxylic cationite, the method comprising the steps of:

- (a) dissolving a monovinyl monomer and a cross-linking agent as co-monomers in a first solvent to form a first mixture;
- (b) adding an initiator of radical polymerization to said first mixture to form a pre-polymer; and

- (c) dispersing said pre-polymer in an aqueous solution of an inorganic salt to form the particulate matter.

36. The method of claim 35, wherein said first solvent is selected from the group consisting of a mixture of an organic solvent and acetic acid, and an aqueous solution of acetic acid.

37. The method of claim 36, wherein said aqueous solution of acetic acid includes an amount of acetic acid in a range of from about 5 percent to about 15 percent volume per volume.

38. The method of claim 36, wherein said aqueous solution of acetic acid includes an amount of acetic acid in a range of from about 60 percent to about 100 percent volume per volume.

39. The method of claim 36, wherein an amount of said monovinyl monomer and an amount of said cross-linking agent are together in a range of from about 50 percent to about 70 percent weight per weight of said first mixture.

40. The method of claim 39, further comprising the step of:

- (d) dispersing the particulate matter in a solution of alcohol.

41. A method for producing a carboxyl cationite, the method consisting essentially of the steps of:

- (a) dissolving a monovinyl monomer and a cross-linking agent as co-monomers in a first solvent to form a first mixture; and
- (b) adding an initiator of radical polymerization to said first mixture to form a swollen block of the carboxyl cationite.

42. The method of claim 41, wherein said first solvent is selected from the group consisting of a mixture of an organic solvent and water, a mixture of an organic solvent and

acetic acid, and an aqueous solution of acetic acid wherein an amount of said acetic acid is in a range of from about 60% to about 100%, volume per volume.

43. The method of claim 42, further comprising the step of:
  - (c) crushing said swollen block to form particulate matter.
44. The method of claim 43, further comprising the step of:
  - (d) washing said particulate matter.

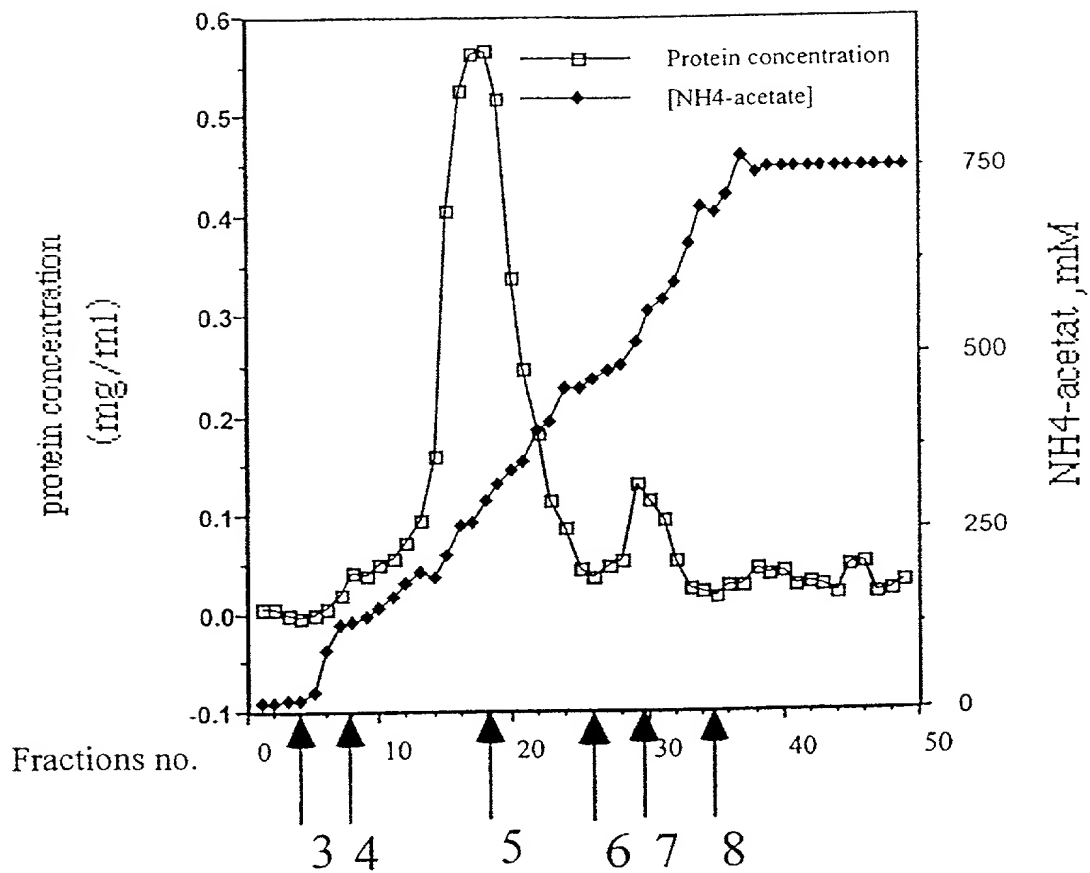


Figure 1A

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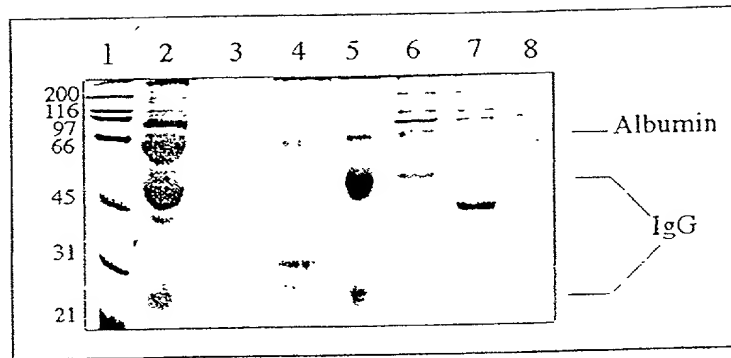


Figure 1B

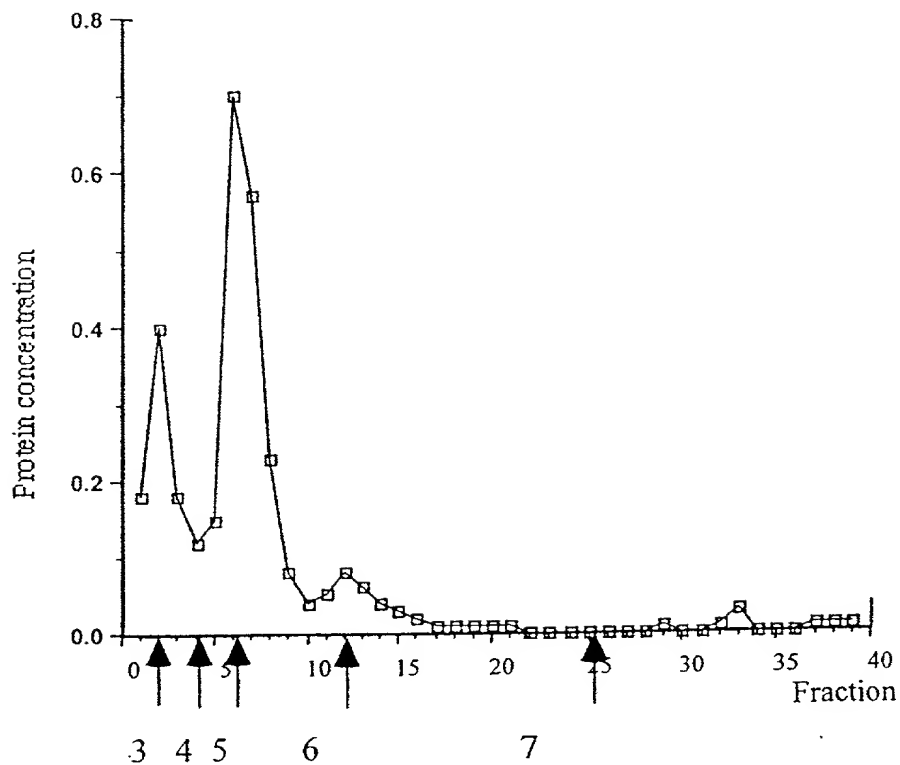


Figure 2A

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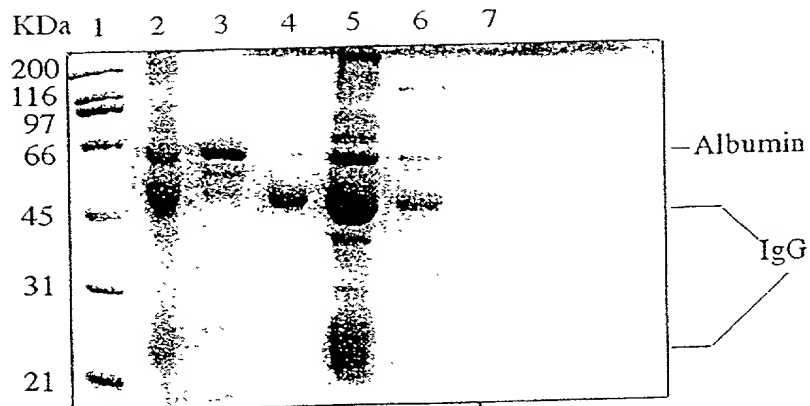


Figure 2B

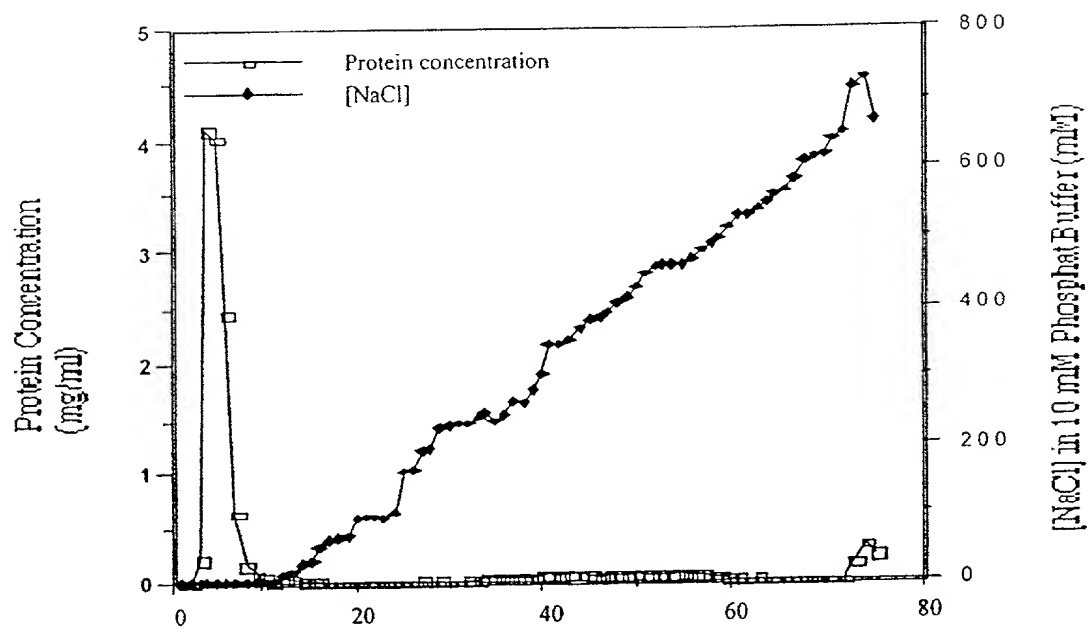


Figure 3A



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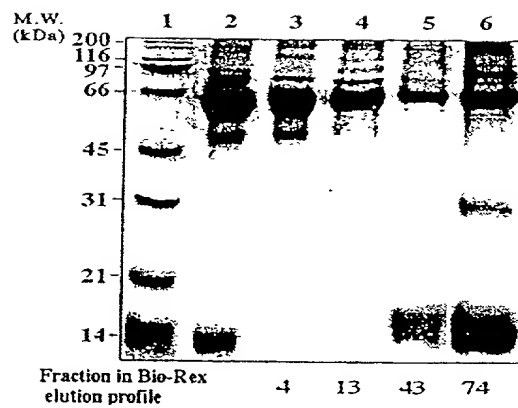


Figure 3B

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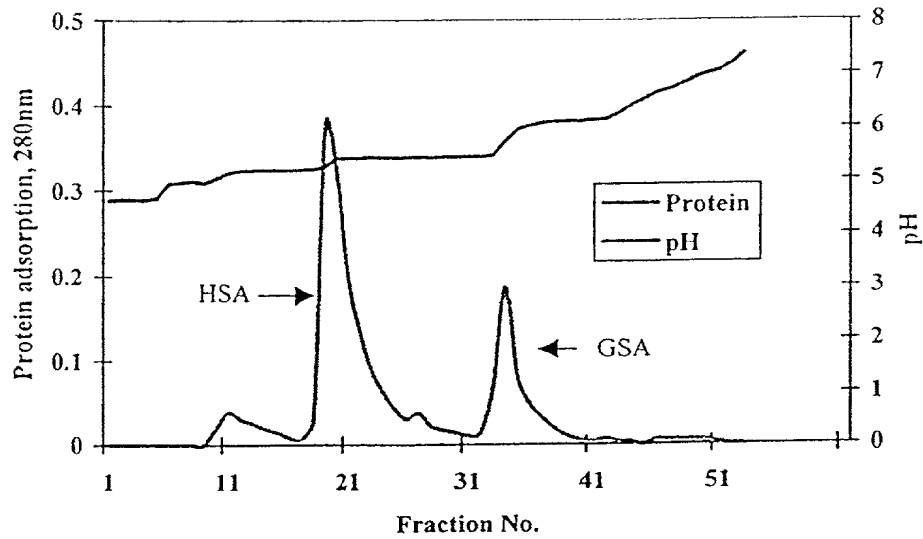


Figure 4

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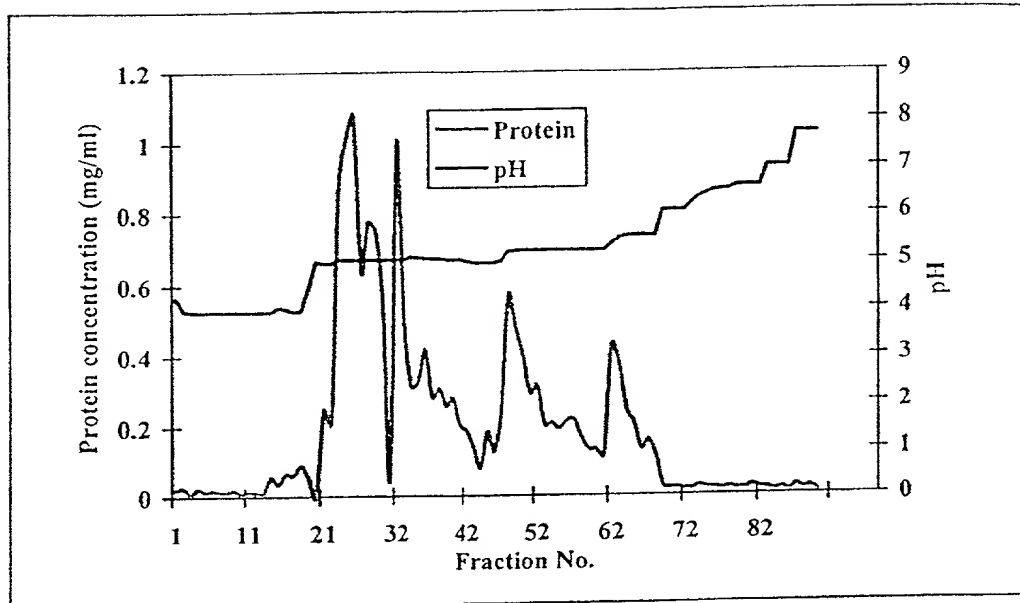


Figure 5

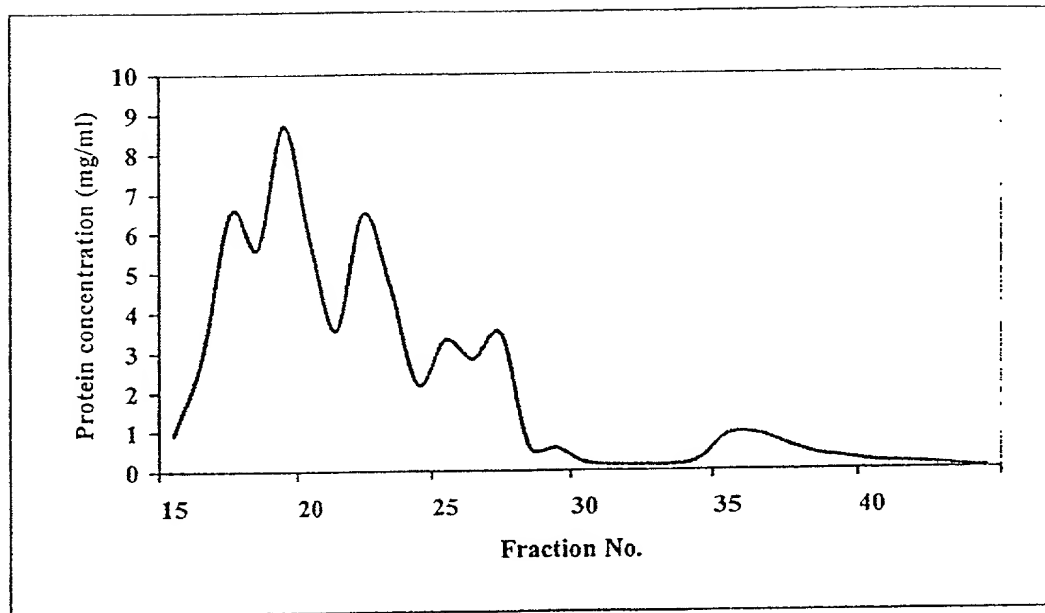


Figure 6

Attorney Docket: 1659/3  
page 1 of 2

Combined Declaration For Patent Application and Power of Attorney

As a below named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name;

I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled CARBOXYLIC CATIONITES AND METHODS OF MANUFACTURE, the specification of which (check one) ☐ is attached hereto.

☒ was filed on 7-Nov-00 as Application Serial No. 09/674,825 and was amended on . I hereby state that I have reviewed and understand the contents of the above identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose information which is material to the patentability of this application in accordance with Title 37, Code of Federal Regulations, § 1.56(a).

I hereby claim foreign priority benefits under Title 35, United States Code, § 119, 365 or 371 of any foreign application(s) for patent or inventor's certificate listed below and have also identified below any foreign application for patent or inventor's certificate having filing date before that of the application on which priority is claimed:

| Prior Foreign Application(s) |                   |                          | Priority Claimed                    |                          |
|------------------------------|-------------------|--------------------------|-------------------------------------|--------------------------|
| <u>IL99/00241</u>            | <u>PCT</u>        | <u>7-May-99</u>          | <input checked="" type="checkbox"/> | <input type="checkbox"/> |
| (number)                     | (Country)         | (Day, Month, Year Filed) | Yes                                 | No                       |
| <u>          </u>            | <u>          </u> | <u>          </u>        | <input type="checkbox"/>            | <input type="checkbox"/> |
| (number)                     | (Country)         | (Day, Month, Year Filed) | Yes                                 | No                       |
| <u>          </u>            | <u>          </u> | <u>          </u>        | <input type="checkbox"/>            | <input type="checkbox"/> |
| (number)                     | (Country)         | (Day, Month, Year Filed) | Yes                                 | No                       |

I hereby claim the benefit under Title 35, United States Code, § 120 of any United States Application(s) listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States application in the manner provided by the first paragraph of Title 35, United States code, § 112, I acknowledge the duty to disclose material information as defined in Title 37, Code of Federal Regulations, § 1.56(a) which occurred between the filing date of the prior application and the national or PCT international filing date of this application:

|                          |                   |                                |
|--------------------------|-------------------|--------------------------------|
| <u>60/084,731</u>        | <u>8-May-98</u>   | <u>pending</u>                 |
| (Application Serial No.) | (Filing Date)     | Status                         |
|                          |                   | (patented, pending, abandoned) |
| <u>          </u>        | <u>          </u> | <u>          </u>              |
| (Application Serial No.) | (Filing Date)     | Status                         |
|                          |                   | (patented, pending, abandoned) |

I hereby appoint the following attorneys, with full power of substitution, association, and revocation, to prosecute this application and to transact all business in the Patent and Trademark Office connected therewith.

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Attorney Docket: 1659/3  
page 1 of 2

### Combined Declaration For Patent Application and Power of Attorney

As a below named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name;

I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled CARBOXYLIC CATIONITES AND METHODS OF MANUFACTURE, the specification of which (check one) ☒ is attached hereto.

☐ was filed on \_\_\_\_\_ as Application Serial No. \_\_\_\_\_ and was amended on \_\_\_\_\_. I hereby state that I have reviewed and understand the contents of the above identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose information which is material to the patentability of this application in accordance with Title 37, Code of Federal Regulations, § 1.56(a).

I hereby claim foreign priority benefits under Title 35, United States Code, § 119 of any foreign application(s) for patent or inventor's certificate listed below and have also identified below any foreign application for patent or inventor's certificate having filing date before that of the application on which priority is claimed:

| Prior Foreign Application(s) |                   |                          | Priority Claimed                        |                             |
|------------------------------|-------------------|--------------------------|---|-----------------------------|
| <u>IL 99/00241</u>           | <u>PCT</u>        | <u>7-May-99</u>          | <input checked="" type="checkbox"/> Yes | <input type="checkbox"/> No |
| (number)                     | (Country)         | (Day, Month, Year Filed) |   |                             |
| <u>          </u>            | <u>          </u> | <u>          </u>        | <input type="checkbox"/> Yes            | <input type="checkbox"/> No |
| (number)                     | (Country)         | (Day, Month, Year Filed) |   |                             |
| <u>          </u>            | <u>          </u> | <u>          </u>        | <input type="checkbox"/> Yes            | <input type="checkbox"/> No |
| (number)                     | (Country)         | (Day, Month, Year Filed) |   |                             |

I hereby claim the benefit under Title 35, United States Code, § 120 of any United States Application(s) listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States application in the manner provided by the first paragraph of Title 35, United States Code, § 112, I acknowledge the duty to disclose material information as defined in Title 37, Code of Federal Regulations, § 1.56(a) which occurred between the filing date of the prior application and the national or PCT international filing date of this application:

|                         |                   |                                |
|-------------------------|-------------------|--------------------------------|
| <u>60/084,731</u>       | <u>8 May-98</u>   | <u>pending</u>                 |
| Application Serial No.) | (Filing Date)     | Status                         |
|                         |                   | (patented, pending, abandoned) |
| <u>          </u>       | <u>          </u> | <u>          </u>              |
| Application Serial No.) | (Filing Date)     | Status                         |
|                         |                   | (patented, pending, abandoned) |

I hereby appoint the following attorneys, with full power of substitution, association, and revocation, to prosecute this application and to transact all business in the Patent and Trademark Office connected therewith.

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Attorney Docket: 1659/3  
page 2 of 2

## Continuation of Combined Declaration For Patent Application and Power of Attorney

I hereby further declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statement may jeopardize the validity of the application of any patent issued thereon.

|   |                         |              |
|---|-------------------------|--------------|
| *FULL NAME OF SOLE OR FIRST INVENTOR    | INVENTOR'S SIGNATURE    | DATE         |
| KARINA ROZHETSKY                        | <i>Karina Rozhetsky</i> | Dec 12, 2000 |
| RESIDENCE                               | CITIZENSHIP             |              |
| PEER STREET 32, 32492 HAIFA, ISRAEL ILX | ISRAELI                 |              |
| POST OFFICE ADDRESS                     |                         |              |
| PEER STREET 32, 32492 HAIFA, ISRAEL     |                         |              |

|                               |                      |      |
|-------------------------------|----------------------|------|
| *FULL NAME OF SECOND INVENTOR | INVENTOR'S SIGNATURE | DATE |
|                               |                      |      |
| RESIDENCE                     | CITIZENSHIP          |      |
|                               | ISRAELI              |      |
| POST OFFICE ADDRESS           |                      |      |
|                               |                      |      |

|                              |                      |      |
|------------------------------|----------------------|------|
| *FULL NAME OF THIRD INVENTOR | INVENTOR'S SIGNATURE | DATE |
|                              |                      |      |
| RESIDENCE                    | CITIZENSHIP          |      |
|                              | ISRAELI              |      |
| POST OFFICE ADDRESS          |                      |      |
|                              |                      |      |

|                               |                      |      |
|-------------------------------|----------------------|------|
| *FULL NAME OF FOURTH INVENTOR | INVENTOR'S SIGNATURE | DATE |
|                               |                      |      |
| RESIDENCE                     | CITIZENSHIP          |      |
|                               | ISRAELI              |      |
| POST OFFICE ADDRESS           |                      |      |
|                               |                      |      |

|                              |                      |      |
|------------------------------|----------------------|------|
| *FULL NAME OF FIFTH INVENTOR | INVENTOR'S SIGNATURE | DATE |
|                              |                      |      |
| RESIDENCE                    | CITIZENSHIP          |      |
|                              | ISRAELI              |      |
| POST OFFICE ADDRESS          |                      |      |
|                              |                      |      |

|                              |                      |      |
|------------------------------|----------------------|------|
| *FULL NAME OF SIXTH INVENTOR | INVENTOR'S SIGNATURE | DATE |
|                              |                      |      |
| RESIDENCE                    | CITIZENSHIP          |      |
|                              | ISRAELI              |      |
| POST OFFICE ADDRESS          |                      |      |
|                              |                      |      |

|                                |                      |      |
|--------------------------------|----------------------|------|
| *FULL NAME OF SEVENTH INVENTOR | INVENTOR'S SIGNATURE | DATE |
|                                |                      |      |
| RESIDENCE                      | CITIZENSHIP          |      |
|                                | ISRAELI              |      |
| POST OFFICE ADDRESS            |                      |      |
|                                |                      |      |

Attorney Docket: 1658/3  
page 2 of 2

## Continuation of Combined Declaration For Patent Application and Power of Attorney

I hereby further declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statement may jeopardize the validity of the application of any patent issued thereon.

|  |  |                 |
|--|--|-----------------|
| *FULL NAME OF SOLE OR FIRST INVENTOR<br>KARINA EHZHETSKY   | INVENTOR'S SIGNATURE<br><i>Karina Ezhetsky</i> | DATE<br>3.11.00 |
| RESIDENCE<br>PEER STREET 32, 32492 HAIFA, ISRAEL IL X      | CITIZENSHIP<br>ISRAELI                         |                 |
| POST OFFICE ADDRESS<br>PEER STREET 32, 32492 HAIFA, ISRAEL |  |                 |

|                               |                        |      |
|-------------------------------|------------------------|------|
| *FULL NAME OF SECOND INVENTOR | INVENTOR'S SIGNATURE   | DATE |
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| POST OFFICE ADDRESS           |                        |      |

|                              |                        |      |
|------------------------------|------------------------|------|
| *FULL NAME OF THIRD INVENTOR | INVENTOR'S SIGNATURE   | DATE |
| RESIDENCE                    | CITIZENSHIP<br>ISRAELI |      |
| POST OFFICE ADDRESS          |                        |      |

|                               |                        |      |
|-------------------------------|------------------------|------|
| *FULL NAME OF FOURTH INVENTOR | INVENTOR'S SIGNATURE   | DATE |
| RESIDENCE                     | CITIZENSHIP<br>ISRAELI |      |
| POST OFFICE ADDRESS           |                        |      |

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|------------------------------|------------------------|------|
| *FULL NAME OF FIFTH INVENTOR | INVENTOR'S SIGNATURE   | DATE |
| RESIDENCE                    | CITIZENSHIP<br>ISRAELI |      |
| POST OFFICE ADDRESS          |                        |      |

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|------------------------------|------------------------|------|
| *FULL NAME OF SIXTH INVENTOR | INVENTOR'S SIGNATURE   | DATE |
| RESIDENCE                    | CITIZENSHIP<br>ISRAELI |      |
| POST OFFICE ADDRESS          |                        |      |

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|--------------------------------|------------------------|------|
| *FULL NAME OF SEVENTH INVENTOR | INVENTOR'S SIGNATURE   | DATE |
| RESIDENCE                      | CITIZENSHIP<br>ISRAELI |      |
| POST OFFICE ADDRESS            |                        |      |